



RESIDUAL DOSES AND SENSITIVITY CHANGE OF POST IR IRSL SIGNALS FROM POTASSIUM FELDSPAR UNDER DIFFERENT BLEACHING CONDITIONS

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Abstract: The residual doses and sensitivity change for potassium-rich feldspar (K-feldspar) have been studied using the post-infrared stimulated luminescence (pIRIR) and multi-elevated-temperature post-IR IRSL (MET-pIRIR) protocols. Laboratory simulated poorly-bleached and well-bleached samples were those K-feldspar grains bleached using a solar simulator for 10 minutes and 8 hours, respectively. The residual doses rise with stimulation temperature and time. The poorly-bleached sample has larger residual doses than the well-bleached sample, especially at high stimulation temperatures. The high-temperature pIRIR signals contain a large amount of hard-to-bleach signals. A decrease of luminescence sensitivity was observed after conducting a high-temperature-treatment in the measurement cycles. The sensitivity decreases significantly between the first and the second cycle. The extent of decrease in sensitivity shows a clear temperature trend. The higher the stimulation temperature of pIRIR signals is, the larger the sensitivity decreases. This decrease is more severe for the poorly-bleached sample than for the well-bleached sample, and could possibly lead to problems in sensitivity correction.

Keywords: residual doses, sensitivity change, infrared stimulated luminescence, K-feldspar.

1. INTRODUCTION

Optically stimulated luminescence (OSL) dating technique has undergone extensive development since the single-aliquot regenerative-dose (SAR) protocol was proposed (Murray and Wintle, 2000; Wintle and Murray, 2006). In the SAR protocol, sensitivity changes caused by laboratory measurements of quartz OSL signals have been successfully monitored and corrected for by measuring the test dose-induced luminescence. The SAR protocol has also been applied to infrared stimulated luminescence (IRSL) signals from feldspar (Wallinga *et al.*, 2000). However, the anomalous fading of IRSL signals from feldspar has long been the major difficulty that hindered its usage for dating of sediments (Wintle, 1973; Spooner, 1994; Huntley and Lamothe, 2001).

It has been reported that following a low-temperature (50°C) IRSL measurement, post-IR IRSL signals (pIRIR) measured at elevated temperatures (>200°C) has a lower fading rate compared to conventional IRSL (Thomsen *et al.*, 2008). This has led to the development of the two-step post-IR IRSL protocol (Buylaert *et al.*, 2009; Thiel *et al.*, 2011). Meanwhile, Li and Li (2011) reported a multi-elevated-temperature post-IR IRSL (MET-pIRIR)

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protocol in which the non-fading component from K-feldspar pIRIR signals can be identified by measuring pIRIR at progressively increased stimulation temperatures (from 50 to 250°C). Some tests based on the MET-pIRIR were also reported using typical aeolian sediment from Loess Plateau and deserts in northern China (e.g. Fu *et al.*, 2012; Li and Li, 2012b).

In these SAR procedures, the reliability of sensitivity correction for these pIRIR signals is the basis for accurate D_e measurements. Previous studies mainly focus on well-bleached sediments, e.g. aeolian loess and desert dune sand. The performance of these protocols for poorly-bleached samples is relatively not well explored, with special regards to sensitivity changes for pIRIR signals from K-feldspar when high-temperature-treatment is adopted. For bleachability of high-temperature pIRIR signals, Li and Li (2011) measured residual doses of high-temperature pIRIR signals for typical aeolian sediments, and reported a dose of a few Gy. Buylaert *et al.* (2012) suggested the high-temperature pIRIR signals have slower bleaching rates under daylight conditions that could cause high residual doses, which may lead to overestimated D_e values for young samples. Some recent studies have successfully tested the performance of SAR pIRIR protocols using small aliquots and single grain technique on young coastal sediments (Reimann *et al.*, 2011, 2012). Using single grain analysis of a natural poorly-bleached sample, Reimann *et al.* (2012) could effectively overcome age overestimation, which are caused by averaging effects in multiple-grain single-aliquot measurements. Studies on the bleachability of pIRIR signals at elevated temperature are remaining to be further studied for various samples to estimate the residual doses.

In this study, we aim to measure the residual doses and the bleachability of pIRIR signals stimulated at various temperatures by using laboratory simulated well- and poorly- bleached K-feldspar samples and by applying the pIRIR and MET-pIRIR protocols. The performance of the SAR sensitivity correction was investigated. Sensitivity changes as function of measurement cycles in these protocols were also investigated to test the effectiveness of the sensitivity correction.

2. SAMPLES AND EXPERIMENTAL DETAILS

A Holocene lakeshore sandy sediment sample from Tibet was prepared using routine procedures, under subdued red light condition (Aitken, 1998). It was firstly treated with 10% HCl acid and 20% H₂O₂ solution to remove carbonates and organic materials, respectively. After conducting dry sieving, K-feldspar grains (125-150 µm) were separated using sodium polytungstate heavy liquid with densities smaller than 2.58 g/cm³. The separated K-feldspar grains were etched with 10% HF acid for 40 min to remove the outer surface. The K-feldspar grains were dried and mounted on 10-mm-diameter aluminium discs

for measurement, using silicone oil as an adhesive. In our experiments, the samples were prepared as medium size aliquots (containing hundreds of grains). An automated Risø TL-DA-20 reader was used for IRSL measurements in the Luminescence Dating Laboratory of The University of Hong Kong. It is equipped with IR diodes (870±40 nm) for stimulation with a power of ~135 mW/cm² at sample position (Bøtter-Jensen *et al.*, 2000). A ⁹⁰Sr/⁹⁰Y beta source was attached to the reader for irradiation with a dose rate of 0.126 Gy/s calibrated on aluminium discs. IRSL signals were detected through a combination of Schott BG-39 and Corning 7-59 filters with the transmission in the range of 320-480 nm.

To simulate poorly-bleached and well-bleached samples, the K-feldspar grains were firstly bleached by an Oriel solar simulator (model: 68820) for 4 hours to reset the natural signal. They were then given a beta dose of 500 Gy. After that, well-bleached and poorly-bleached samples were prepared as:

- 1) Well-bleached sample (WB): this sample was bleached under the solar simulator for 8 hours.
- 2) Poorly-bleached sample (PB): this sample was bleached by solar simulator for only 10 minutes. It should be noted that PB in this study is homogeneously poorly bleached, while in nature poorly-bleached samples mostly refer to heterogeneously poorly bleached.

3. MEASURING RESIDUAL DOSES

To evaluate the bleachability for pIRIR signals that stimulated at various temperatures, the residual doses for pIRIR signals were measured using the MET-pIRIR (50, 100, 150, 200, 250, 300°C) (Li and Li, 2012b), pIRIR (50, 225°C), pIRIR (50, 290°C) and pIRIR (200, 290°C) protocols (Buylaert *et al.*, 2009; Thiel *et al.*, 2011; Li and Li, 2012a). The details of these protocols are shown in **Table 1**.

The residual doses of the two-step pIRIR protocols range from 2.2 to 29.0 Gy for PB, 0.6 to 7.2 Gy for WB; while the results for MET-pIRIR range from 7.7 to 49.0 Gy for PB and 4.1 to 20.4 Gy for WB (**Table 2**). Results have also been reported that a variety of residual doses up to ~55 Gy were found in high-temperature pIRIR signals from typical aeolian sediments from Eurasia (Li *et al.*, 2013a). The residual doses found in our study are large, even after 8h solar simulator bleaching. A significantly smaller residual dose (less than 1 Gy) has been reported for well-bleached sample after prolonged bleaching in a solar simulator (Reimann *et al.*, 2012) and for a modern natural sample (Madsen *et al.*, 2011), using the pIRIR 180 and 150°C signals, respectively. Under different bleaching conditions including solar simulator, daylight and infrared light source, a similar small residual dose is reported for the well-bleached sample and a dose of 1-3.5 Gy for the poorly-bleached sample (after 10 min IR bleaching) using the pIRIR 150°C signal (Reimann

Table 1. Residual doses of well- and poorly- bleached samples (WB and PB), measured with SAR protocol (Buylaert *et al.*, 2009; Thiel *et al.*, 2011; Li and Li, 2012a, 2012b).

| Step | MET-pIRIR (50, 100, 150, 200, 250, 300°C) | pIRIR (50, 225°C) | pIRIR (50, 290°C) | pIRIR (200, 290°C) |
|------|-------------------------------------------|-------------------------------|-------------------------------|-------------------------------|
| 1 | Dose, D_i^a | Dose, D_i^a | Dose, D_i^a | Dose, D_i^a |
| 2 | Preheat at 320°C, 60 s | Preheat at 250°C, 60 s | Preheat at 320°C, 60 s | Preheat at 320°C, 60 s |
| 3 | IRSL 100 s at 50°C (L_x) | IRSL 100 s at 50°C (L_x) | IRSL 200 s at 50°C (L_x) | IRSL 200 s at 200°C (L_x) |
| 4 | IRSL 100 s at 100°C (L_x) | IRSL 100 s at 225°C (L_x) | IRSL 200 s at 290°C (L_x) | IRSL 200 s at 290°C (L_x) |
| 5 | IRSL 100 s at 150°C (L_x) | Test dose, D_t | Test dose, D_t | Test dose, D_t |
| 6 | IRSL 100 s at 200°C (L_x) | Preheat at 250°C, 60 s | Preheat at 320°C, 60 s | Preheat at 320°C, 60 s |
| 7 | IRSL 100 s at 250°C (L_x) | IRSL 100 s at 50°C (T_x) | IRSL 200 s at 50°C (T_x) | IRSL 200 s at 200°C (T_x) |
| 8 | IRSL 100 s at 300°C (L_x) | IRSL 100 s at 225°C (T_x) | IRSL 200 s at 290°C (T_x) | IRSL 200 s at 290°C (T_x) |
| 9 | Test dose, D_t | IRSL 40 s at 290°C | IRSL 100 s at 325°C | IRSL 40 s at 325°C |
| 10 | Preheat at 320°C, 60 s | Return to step 1 | Return to step 1 | Return to step 1 |
| 11 | IRSL 100 s at 50°C (T_x) | | | |
| 12 | IRSL 100 s at 100°C (T_x) | | | |
| 13 | IRSL 100 s at 150°C (T_x) | | | |
| 14 | IRSL 100 s at 200°C (T_x) | | | |
| 15 | IRSL 100 s at 250°C (T_x) | | | |
| 16 | IRSL 100 s at 300°C (T_x) | | | |
| 17 | IRSL 100 s at 325°C | | | |
| 18 | Return to step 1 | | | |

^a For the 'natural' sample, $i=0$ and $D_0=0$. The whole sequence is repeated for several regenerative doses including a zero dose and a repeated dose.

Table 2. Residual doses for the MET-pIRIR protocol (50, 100, 150, 200, 250, 300°C) and pIR-IRSL protocol of (50, 225°C), (50, 290°C) and (200, 290°C).

| Temperature (°C) | Residual dose for PB (Gy) | Residual dose for WB (Gy) |
|------------------|---------------------------|---------------------------|
| 50 | 7.7 | 4.1 |
| 100 | 21.9 | 11.6 |
| 150 | 27.4 | 12.8 |
| 200 | 30.9 | 13.5 |
| 250 | 34.4 | 15.0 |
| 300 | 49.0 | 20.4 |
| ----- | | |
| 50 | 2.2 | 0.6 |
| 225 | 15.2 | 5.4 |
| ----- | | |
| 50 | 7.8 | 2.4 |
| 290 | 27.6 | 6.6 |
| ----- | | |
| 200 | 20.5 | 5.5 |
| 290 | 29.0 | 7.2 |

Note: the residual dose is obtained using data in the first 4 second for integration. The total stimulation time is 100 s. The residual doses are the average from three aliquots.

and Tsukamoto, 2012). Using the pIRIR (50, 290) protocol, Buylaert *et al.* (2012) reported a linear relationship between residual doses of the pIRIR 290°C signal and burial dose for a set of samples having different burial doses, after 4 hour bleaching under a solar simulator. They suggest an unbleachable residual dose of ~4 Gy. After bleaching a sample for 4 hour in a solar simulator, a comparable residual dose of ~7 Gy has been reported for the pIRIR 290°C signal (Reimann *et al.*, 2011). These

results suggest that the residual doses for different pIRIR signals from various samples could vary significantly.

Fig. 1 shows the residual doses for the MET-pIRIR (50, 100, 150, 200, 250, 300°C) protocol, with the lower series for WB and the higher series for PB, respectively. The residual doses at each stimulation temperature are plotted against the stimulation time. D_e values were calculated at successive intervals of 2 s. Each point is the average from three aliquots. Only the residual doses of the first 20 s were plotted. The total stimulation time is 100 s. It is observed that PB has higher residual doses than WB. The difference between residual doses for PB and WB increases with stimulation temperature. High-temperature pIRIR signals are relatively hard-to-bleach and contain larger residuals. A modified-low-temperature MET-pIRIR protocol has been suggested to avoid large residuals using the stimulation temperatures of 110, 140 and 170°C (Fu and Li, 2013). It is also reported for the two-step pIRIR protocols that lowering temperature for preheat and the second IR stimulation can reduce the residual doses (e.g. Madsen *et al.*, 2011; Reimann *et al.*, 2011). It is noted in Fig. 1 that an abrupt rise of residual dose from 250 to 300°C is observed for both WB and PB, implying that the luminescence characteristics may have a significant change during this temperature interval.

It is also observed that the residual doses of low-temperature pIRIR signals show a rise with stimulation time in D_e -t plot (Fig. 1), indicating that the initial part of signals relate more to easy-to-bleach signals; while the later part relates more to hard-to-bleach signals (Li and Wintle, 1992). For PB, this increment in residual doses is much smaller for 100°C stimulation than 50°C, implying that the proportion for easy-to-bleach signals for 100°C stimulation decreases significantly compared to that of

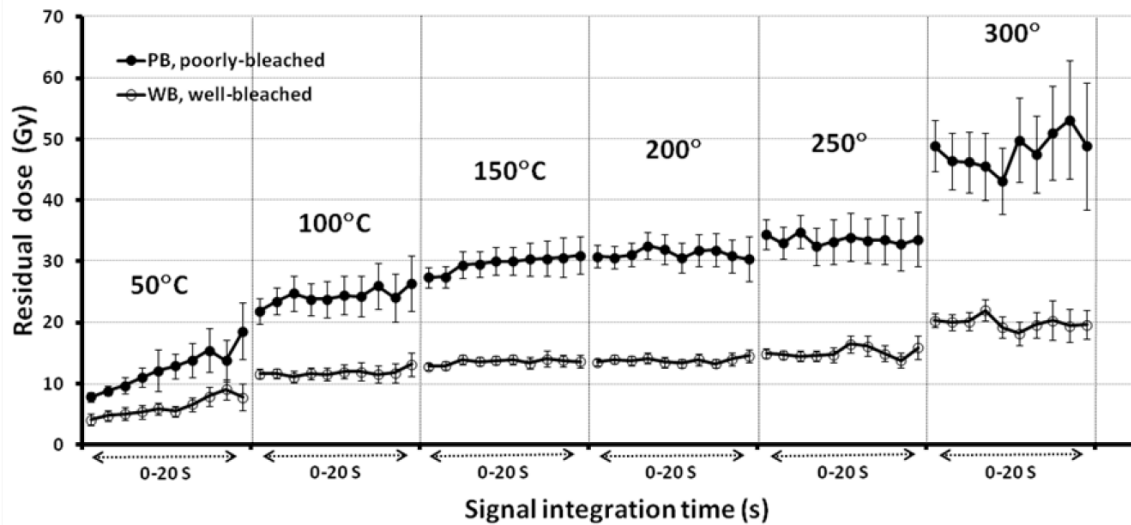


Fig. 1. Residual dose for well-bleached sample (WB, the lower series) and poorly-bleached sample (PB, the upper series) changes with the stimulation time and temperatures, from pIRIR signals that were stimulated at various temperatures, using MET-pIRIR protocol of (50, 100, 150, 200, 250, 300°C) (Li and Li, 2012b). D_e values were calculated at successive intervals of 2 second. Only the residual doses of first 20 s were plotted. The total stimulation time is 100 s. Each point is the average from three aliquots, so as results below. The details of MET-pIRIR protocol (50, 100, 150, 200, 250, 300°C) are shown in Table 1.

the 50°C signals. The residual dose gradually reaches a plateau at 150°C or higher, implying that after this temperature luminescence associated with hard-to-bleach signals dominates the total signal. For WB, similar pattern is observed but the increment in residual doses is much smaller in the 50°C signals. The D_e -t plateau occurs at a relatively low temperature (100°C). PB shows larger increment of residual doses with stimulation time, especially for low-temperature signals. This suggests that major part of residual low-temperature pIRIR signal for PB is from the easy-to-bleach signals. These easy-to-bleach signals remain large in PB, but can be bleached under prolonged solar bleaching in WB, because WB shows smaller increment of residual doses with stimulation time.

Fig. 2 shows the residual doses for pIRIR protocols of (50, 225°C), (50, 290°C) and (200, 290°C), for both PB (upper series) and WB (lower series). Each point is the average from three aliquots. Similar patterns to the MET-pIRIR results can be observed for the two-step pIRIR protocols. PB has larger residual doses than WB. A rise in D_e -t plot is found for low-temperature IRSL signals while high-temperature pIRIR shows a negligible D_e -t increment. It is noted that in the pIRIR protocol (50, 290°C), the 290°C-pIRIR also shows an increasing D_e -t (PB in Fig. 2b). This suggests that after the first step IRSL (50°C), easy-to-bleach signals are still present in the following 290°C pIRIR signals. However, in the pIRIR protocols of (50, 225°C) and (200, 290°C), a smaller

increment of D_e -t for high-temperature pIRIR signals is observed. This different trend of D_e -t for high-temperature pIRIR signals implies that the bleaching rate of pIRIR signals of different stimulation temperatures varies significantly. Different combination of the prior-IR and post-IR stimulation temperatures can result in different proportions between the easy-to-bleach and hard-to-bleach components. Therefore, different residual doses are expected.

4. MEASURING SENSITIVITY CHANGES

In order to study the sensitivity changes of the pIRIR signals as function of measurement cycles, aliquots of WB and PB were given a fixed dose of 25 Gy as regenerative dose (L_x) and a test dose of 6.3 Gy was given prior to the T_x measurement, repeated for four to six cycles, using different pIRIR protocols: MET-pIRIR protocol with IR stimulation temperatures of 50, 100, 150, 200, 250°C (Li and Li, 2011) and a simplified MET-pIRIR protocol with IR stimulation temperature at 110, 140 and 170°C (Fu and Li, 2013), and two-step pIRIR protocols (pIRIR (50, 225°C) and pIRIR (50, 290°C)) (Buylaert *et al.* 2009; Thiel *et al.*, 2011). The details of these protocols are shown in Table 3. Six aliquots of WB and PB were used for each experiment. Three of the six aliquots were exposed to a hot bleach (high-temperature IRSL stimulation, Table 3) after each T_x measurement at the end of each SAR cycle. The other three aliquots were not exposed to a hot bleach.

Table 3. Sensitivity change as function of measurement cycles, measured with SAR protocols (Buylaert *et al.*, 2009; Thiel *et al.*, 2011; Li and Li, 2011; Fu and Li, 2013).

| Step | MET-pIRIR (110, 140, 170°C) | MET-pIRIR (50, 100, 150, 200, 250°C) | pIRIR (50, 225°C) | pIRIR (50, 290°C) |
|------|-------------------------------|--------------------------------------|-------------------------------|-------------------------------|
| 1 | Dose, 25 Gy | Dose, 25 Gy | Dose, 25 Gy | Dose, 25 Gy |
| 2 | Preheat at 200°C, 60 s | Preheat at 300°C, 10 s | Preheat at 250°C, 60 s | Preheat at 320°C, 60 s |
| 3 | IRSL 100 s at 110°C (L_x) | IRSL 100 s at 50°C (L_x) | IRSL 100 s at 50°C (L_x) | IRSL 200 s at 50°C (L_x) |
| 4 | IRSL 100 s at 140°C (L_x) | IRSL 100 s at 100°C (L_x) | IRSL 100 s at 225°C (L_x) | IRSL 200 s at 290°C (L_x) |
| 5 | IRSL 100 s at 170°C (L_x) | IRSL 100 s at 150°C (L_x) | Test dose, 6.3 Gy | Test dose, 6.3 Gy |
| 6 | Test dose, 6.3 Gy | IRSL 100 s at 200°C (L_x) | Preheat at 250°C, 60 s | Preheat at 320°C, 60 s |
| 7 | Preheat at 200°C, 60 s | IRSL 100 s at 250°C (L_x) | IRSL 100 s at 50°C (T_x) | IRSL 200 s at 50°C (L_x) |
| 8 | IRSL 100 s at 110°C (T_x) | Test dose, 6.3 Gy | IRSL 100 s at 225°C (T_x) | IRSL 200 s at 290°C (L_x) |
| 9 | IRSL 100 s at 140°C (T_x) | Preheat at 300°C, 10 s | IRSL 40 s at 290°C | IRSL 100 s at 325°C |
| 10 | IRSL 100 s at 170°C (T_x) | IRSL 100 s at 50°C (T_x) | Return to step 1 | Return to step 1 |
| 11 | IRSL 100 s at 280°C | IRSL 100 s at 100°C (T_x) | | |
| 12 | Return to step 1 | IRSL 100 s at 150°C (T_x) | | |
| 13 | | IRSL 100 s at 200°C (T_x) | | |
| 14 | | IRSL 100 s at 250°C (T_x) | | |
| 15 | | IRSL 100 s at 320°C | | |
| 18 | | Return to step 1 | | |

Note: the sensitivity change is measured with a fixed regenerative dose of 25 Gy and test dose of 6.3 Gy, repeated for 4 to 6 times for both WB and PB.

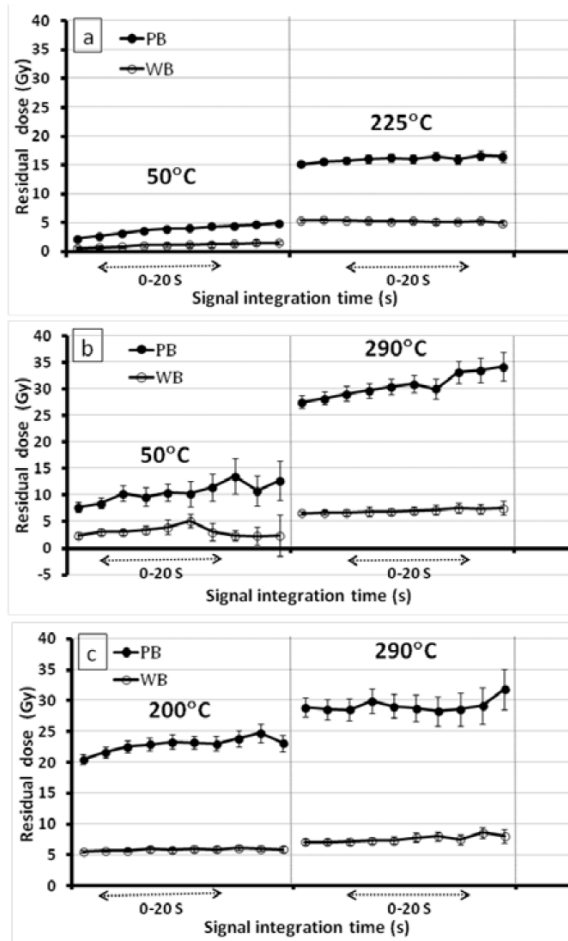


Fig. 2. Residual dose for the pIR-IRSL protocols of WB and PB for: a) 50, 225°C (Buylaert *et al.*, 2009); b) 50, 290°C (Thiel *et al.*, 2011); c) 200, 290°C (Li and Li, 2012a). The legend and integration of signals is same to that of Fig. 1 with the measuring method changed to two-step pIRIR protocols. The details of pIRIR protocols are shown in Table 1.

Sensitivity change in the MET-pIRIR protocol

MET-pIRIR protocol (110, 140, 170°C)

The MET-pIRIR protocol (110, 140 and 170°C) (Fu and Li, 2013) was repeated for 6 cycles for WB and PB. Fig. 3 shows the normalized L_x (bold line, left scale), T_x (dashed line, left scale) and L_x/T_x (scattered data points in lower part of figure, right scale) as a function of measurement cycles at 110 (blue), 140 (green) and 170°C (red). Figs. 3a and b show the results for PB and WB, respectively, without a hot bleach. Figs. 3c and d show the results for PB and WB with a hot bleach (IRSL 280°C, 100 s) at the end of each cycle.

For PB, the L_x 110°C (blue bold line) rises with cycle number (Fig. 3a). L_x 140°C (green bold) and L_x 170°C (red bold) decrease from the first cycle to the second cycle. This is expected because the high-temperature pIRIR signals have more residual doses than the low temperature IRSL signals. This suggests that after the short solar simulator bleaching (10 min), the 110°C IRSL signals can be effectively bleached while a considerable amount of residual signals remained in high-temperature pIRIR signals. In contrast, all T_x (dashed lines) increase with cycle number. The higher the stimulation temperature of pIRIR is, the smaller the sensitivity increases in repeated measurement cycles, indicating variation in sensitivity changes for pIRIR signals stimulated under different temperatures.

For WB, it is observed that L_x (bold line) constantly increases with the stimulation temperature (Fig. 3b). The higher the stimulation temperature of pIRIR is, the less the sensitivity increases. T_x shows a similar pattern as L_x . There is no systematic variation of L_x/T_x with cycle number (Fig. 3b), indicating that the sensitivity change can be corrected for these samples. The results suggest that WB

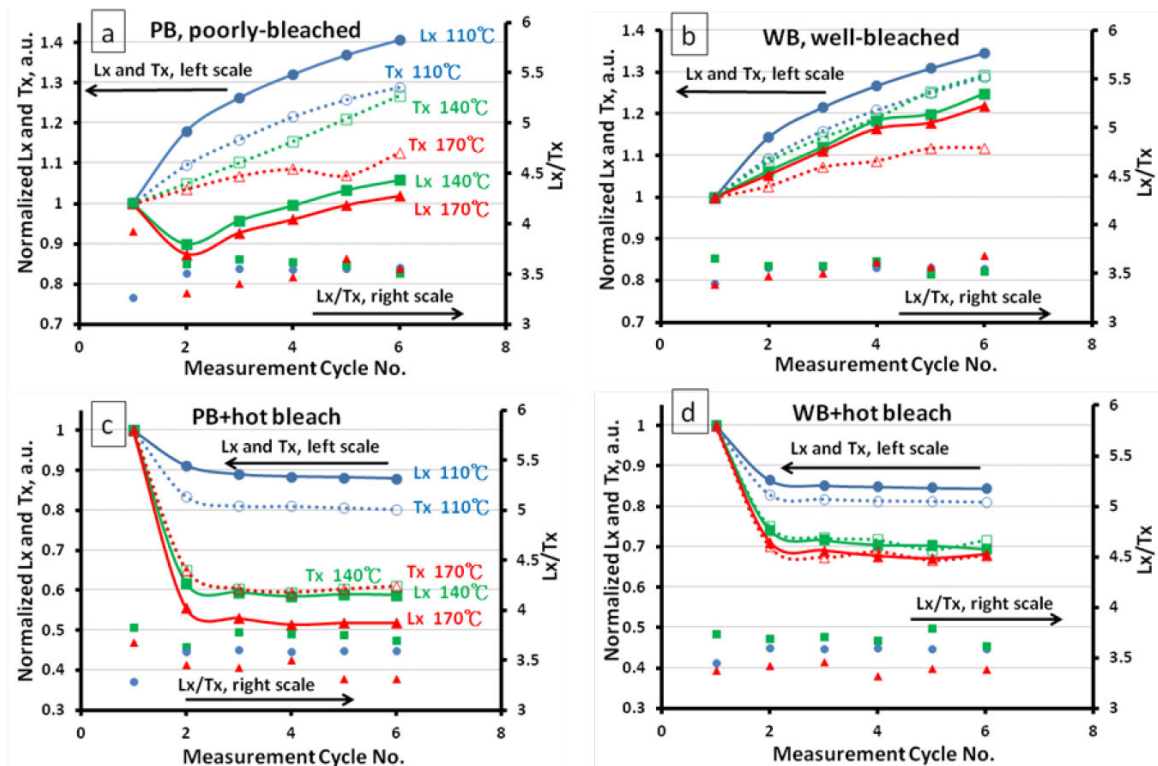


Fig. 3. To monitor sensitivity changes as function of measurement cycles, WB and PB were given a constant dose of 25 Gy for L_x measurement and 6.3 Gy for T_x measurement, using the MET-pIRIR protocol (110, 140 and 170°C), repeated for six cycles (Fu and Li, 2013). Details of the protocol are described in Table 3. Normalized L_x (bold line, left scale), T_x (dashed line, left scale) and L_x/T_x ratio (points, right scale) changes with the measurement cycle numbers for MET-pIRIR protocol, at 110°C (blue), 140°C (green) and 170°C (red). Six aliquots were measured for WB and PB, respectively, and a hot bleach at the end of every measurement cycle (IRSL 280°C, 100 s) was adopted to every three aliquots of the six. a) for poorly-bleached sample PB; b) for well-bleached sample WB; c) for PB+hot bleach; d) for WB+hot bleach.

was well bleached and hence negligible residual signals present in the first L_x measurement, even for the pIRIR 170°C signals.

Figs. 3c and **d** show the results with a hot bleach for PB and WB, respectively. Different patterns from **Figs. 3a** and **b** were observed: both L_x and T_x decrease significantly from first cycle to the second cycle. They keep decreasing continuously in a smaller degree from the second to the third cycles. After the third cycle, the sensitivity generally remains unchanged. Larger decreases are observed for both L_x and T_x for pIRIR signals stimulated at high temperatures. These larger decreases are contributed from both residual and sensitivity decrease. As shown for the L_x 140 and 170°C of PB (**Fig. 3a**), an apparent drop is found from the first to the second cycle, but not in the L_x 110°C, suggesting that considerable amount of residual signal presented in the high-temperature pIRIR signals. However, T_x increases with measurement cycle number in **Figs. 3a** and **b**, while decreases in **Figs. 3c** and **d**. Comparing with **Fig. 3a**, the extent of decrease for 140 and 170°C L_x in **Fig. 3c** is significantly larger after the hot bleach. It is thus suggested that the sensitivity for the pIRIR signals decreases after the hot bleach. WB also shows a similar decrease

(**Fig. 3d**). From the first cycle to the second, the extent of decrease for T_x in **Fig. 3d** is smaller than that in the **Fig. 3c**, indicating that the sensitivity decrease is larger for PB than for WB after the hot bleach. Negligible difference for sensitivity decrease is observed between PB and WB after the first cycle, suggesting that the sensitivity decrease mainly occurs in the initial measurement cycles, especially between the first and the second cycles. Sensitivity change from the measurement of natural signal to immediately following T_x measurement has also been reported for quartz OSL signals (Singhvi *et al.*, 2011).

MET-pIRIR protocol (50, 100, 150, 200, 250°C)

The 50°C interval MET-pIRIR protocol (Li and Li, 2011), was tested using both WB and PB. **Figs. 4a** and **b** show the results for PB and WB without a hot bleach, respectively. Similar to **Fig. 3a**, both L_x and T_x of PB show a general increasing trend between the first and the second cycles for low-temperature pIRIR signals (**Fig. 4a**). However, in contrast to **Fig. 3a** high-temperature pIRIR signals decrease with the cycles (**Fig. 4a**). As discussed above, the residual signal is one of reasons accounting for the L_x decrease between the first cycle and the second cycle. For the T_x signals at high temperatures,

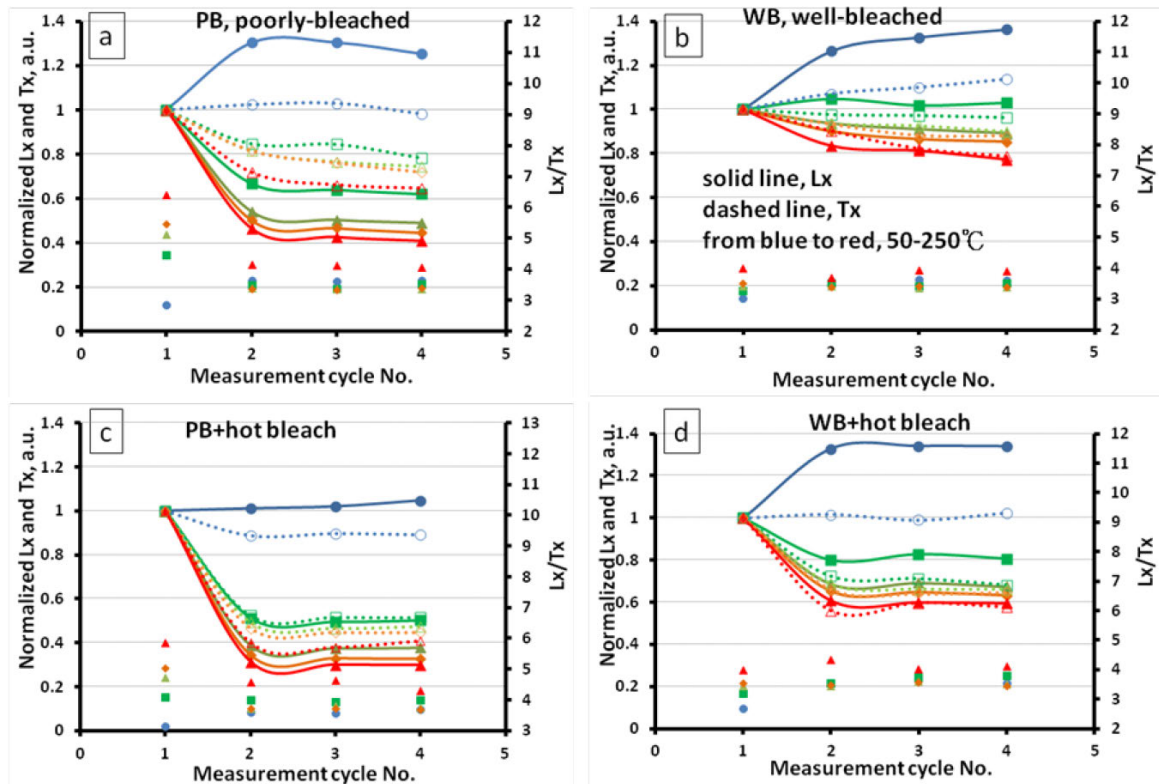


Fig. 4. Sensitivity changes of WB and PB as function of measurement cycles (a constant dose of 25 Gy for L_x measurement and 6.3 Gy for T_x measurement), using the MET-pIRIR protocol (50, 100, 150, 200, 250°C), repeated for four cycles (Li and Li, 2011). Six aliquots were measured for WB and PB, respectively, and a hot bleach at the end of every measurement cycle (IRSL 320°C, 100 s) was adopted to every three of the six aliquots. Normalized L_x (bold line), T_x (dashed line) and L_x/T_x ratio (points) changes with the measurement cycle numbers for MET-pIRIR protocol, at 50°C (blue), 100°C (green), 150°C (yellow-green), 200°C (yellow) and 250°C (red). a) for PB; b) for WB; c) for PB+hot bleach; d) for WB+hot bleach. The experimental conditions are shown in Table 3. Note that the figure contains many lines and using different colours would be easier to distinguish these IRSL signals stimulated at various temperatures.

they also decrease with cycles, suggesting that a sensitivity decrease has already occurred in the MET-pIRIR (50-250°C) signals, even without the hot bleach. T_x and L_x for WB also show such decrease at temperatures above 150°C (Fig. 4b). Comparing with PB (Fig. 4a), the extent of decrease between the first and the second cycle is smaller for WB (Fig. 4b). This smaller extent of decrease for T_x signals for WB suggests that sensitivity decrease is more severe for PB than WB. This behaviour was not observed in MET protocol of (110, 140, 170°C) (Figs. 3a and b). This can be explained as the highest stimulation temperature (170°C) or the preheat (200°C) in the MET (110, 140, 170°C) is not high enough to induce this sensitivity changes. After adopting a hot bleach, sensitivity decrease is observed in Figs. 3c and d, suggesting that high-temperature preheat or pIRIR stimulation in the high temperatures of MET protocol (50-250°C) plays a similar role to the hot bleach in terms of inducing the sensitivity changes.

The results suggest that sensitivity decrease could occur from L_x to the immediately following T_x measurement after conducting high-temperature treatment in the

pIRIR measurement. Since it is observed that sensitivity significantly decreases between the first and the second cycle, e.g. from L_1 to L_2 or from T_1 to T_2 (Figs. 3c and d, and high temperature pIRIR signals in Figs. 4a and b), if high temperature treatment is adopted in pIRIR protocols, it is reasonable to assume that the sensitivity decrease should be the largest between the measurement of L_n and T_n (or L_1 and T_1 in this study). In the following cycles, such decrease still presents but in a smaller degree. In this case, after adopting a high-temperature treatment in the pIRIR procedures, T_x may not be successfully used to correct the sensitivity changes as the preceding high-temperature treatment could induce considerable sensitivity change for the T_x , especially in the first cycle measurement.

Figs. 4c and d show the results with a hot bleach (IRSL 320°C, 100 s) for PB and WB, respectively. Similar patterns to Figs. 3c and d are observed. The extent of sensitivity decrease is larger for high-temperature pIRIR signals, suggesting that sensitivity decrease is more severe for high-temperature pIRIR signals. The T_x decrease is more severe for PB than WB (Fig. 4d), suggesting that

PB has a larger sensitivity decrease after a hot bleach. The sensitivity correction (points in Fig. 4) also presents larger scatter, especially for the first measurement cycle. After the first cycle, negligible difference in sensitivity decrease is observed between WB and PB.

It has been reported that a high-temperature heat treatment (up to 600°C) can reduce the IRSL sensitivity from K-feldspar (e.g. Richardson, 1994; Li *et al.*, 2013b). In this study, sensitivity decrease is observed for MET-pIRIR signals for both WB and PB. It is found to be more severe in our poorly-bleached samples, suggesting that caution should be taken when dating poorly bleached sediments when high-temperature-treatment is adopted. It is noted that the sensitivity decrease mainly occurs in the initial measurement cycles, though our observation of sensitivity change is found from the T_x signals between the first and the second cycles. Based on our observations, the sensitivity decrease mainly occurs in the initial measurement cycles. Hence it is interpreted as considerable sensitivity decrease between the measurement of L_n and T_n could happen after adopting a high-temperature treatment. The situation of sensitivity correction is more severe for PB as it shows larger sensitivity decrease compared with WB. It is thus suggested that caution should be exercised when using high-temperature treatment in

these protocols. Sensitivity changes have been investigated during OSL measurement of natural signal in the quartz SAR procedure (e.g. Murray and Wintle, 2000), and it has been reported that sensitivity correction does not take into account the changes in sensitivity that could happen during the readout of the natural signal (Singhvi *et al.*, 2011). It is noted that the luminescence mechanism for these two minerals is significantly different, but for reliable sensitivity correction to be met in the SAR protocol, possible sensitivity changes between L_n and T_n measurement must be carefully considered for both minerals.

Sensitivity change in the two-step pIR-IRSL protocol

Similar measurement was then carried out using the pIRIR (50, 225°C) and pIRIR (50, 290°C) protocols, for both WB and PB (Buylaert *et al.*, 2009; Thiel, *et al.*, 2011). The results are shown in Figs. 5 and 6. Similar behaviours in the MET-pIRIR results are observed. In the two step-pIRIR results, sensitivity also decreases after the hot bleach. The sensitivity decreases significantly from the first to the second cycle. If high temperature treatment is used in the pIRIR measurement, a large sensitivity decrease could occur between the L_n measurement and the T_n measurement following it. This could lead to prob-

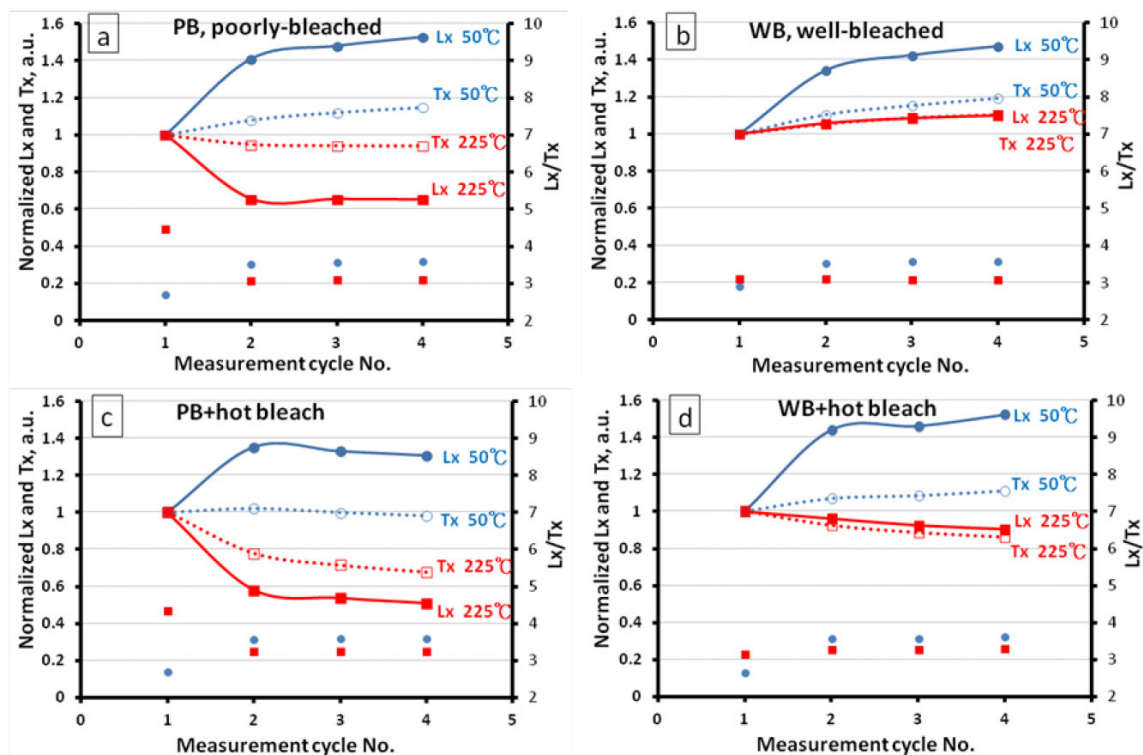


Fig. 5. Sensitivity changes of WB and PB as a function of measurement cycle number, using pIRIR (50, 225°C), repeated for four cycles (Buylaert *et al.*, 2009). Six aliquots were measured for WB and PB, respectively, and a hot bleach at the end of every measurement cycle (IRSL 290°C, 40 s) was applied to every three of the six aliquots. Normalized L_x (bold line), T_x (dashed line) and L_x/T_x ratio (points) changes with the measurement cycle numbers for pIR-IRSL protocol, at 50°C (blue) and 225°C (red). a) for PB; b) for WB; c) for PB+hot bleach; d) for WB+hot bleach. Details of the protocol are shown in Table 3.

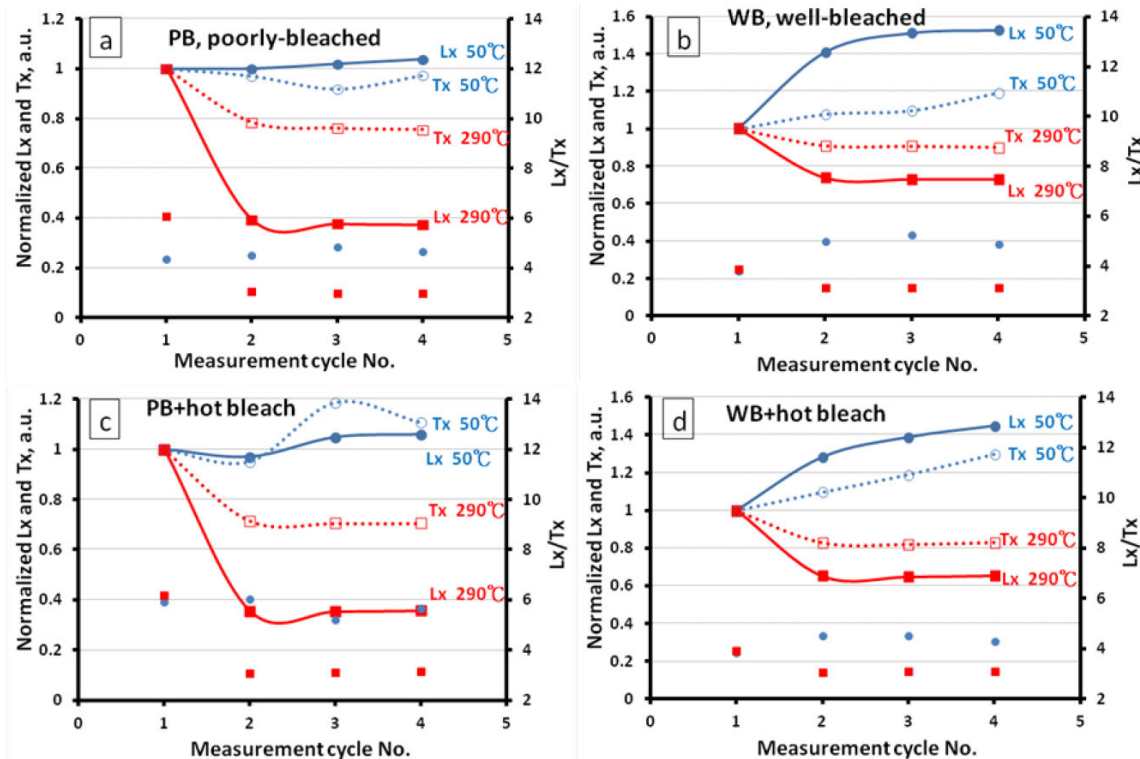


Fig. 6. Sensitivity changes of WB and PB as function of measurement cycles, using pIRIR (50, 290°C), repeated for four cycles (Thiel *et al.*, 2011). Six aliquots were measured for WB and PB, respectively, and a hot bleach at the end of every measurement cycle (IRSL 325°C, 100 s) was adopted to every three of the six aliquots. Normalized L_x (bold line), T_x (dashed line) and L_x/T_x ratio (points) changes with the measurement cycle numbers for pIR-IRSL protocol, at 50°C (blue) and 290°C (red). a) for PB; b) for WB; c) for PB+hot bleach; d) for WB+hot bleach. Similar patterns to Fig. 4 can be observed. Details of the protocol are shown in Table 3.

lems in sensitivity correction as high-temperature treatment during the natural signal measurement could induce severe sensitivity decrease for its following T_n signal. In this case, T_n could not correctly reflect the sensitivity for L_n .

The problem of sensitivity change is more severe for the poorly-bleached sample, because it is found that the extent of sensitivity decrease is larger for PB than for WB. Our result also suggests that the sensitivity decrease is more severe for high-temperature pIRIR signals, as they show larger sensitivity decrease compared with the low-temperature pIRIR (Figs. 5c and d, 6c and d). In this case, larger sensitivity decrease between L_n and T_n measurement occurs and leads to a more severe problem for sensitivity correction. The result suggests that caution must be exercised when using high temperature treatment in these protocols, because the sensitivity decreases by the high temperature treatment and then the sensitivity correction may be problematic. A similar problem has been mentioned by Vasiliniuc *et al.* (2012), who suggested that the pIRIR 300°C signal appears to suffer from dose dependent initial sensitivity changes for their oldest samples. However, several recent works have reported pIRIR protocols generally give OSL ages in agreement with independent age controls (e.g. Buylaert *et al.*, 2012), suggesting that sensitivity-corrections are generally satis-

factory in these studies. This implies that sensitivity-correction problem may not be a common phenomenon for all K-feldspar. Since our observation of sensitivity change is from the T_x signals between different measurement cycles, we suggest that more attention should be paid to sensitivity changes in various pIRIR protocols, as reliable sensitivity correction is the basis for accurate D_e measurements.

5. CONCLUSION

The residual doses for laboratory simulated well- and poorly- bleached samples (WB and PB) were measured using the two-step pIR-IRSL protocols of (50, 225°C), (50, 290°C) and (200, 290°C), and MET-pIRIR protocol (50, 100, 150, 200, 250, 300°C). The residual doses range from 2.2 to 49.0 Gy for PB and 0.6 to 20.4 Gy for WB. The results suggest that PB has larger residual dose than WB, especially for the high-temperature pIRIR signals. The D_e -t plots reach plateaus for the high-temperature pIRIR signals, for both the pIR-IRSL and MET-pIRIR protocols.

Sensitivity change as a function of measurement cycles for WB and PB were investigated using the two-step pIRIR protocols of (50, 225°C) and (50, 290°C), and

MET-pIRIR protocols of (50, 100, 150, 200, 250°C) and (110, 140, 170°C). Sensitivity decrease is observed after a high-temperature treatment is adopted, especially between the first and second cycles. This decrease is more severe for PB and more severe for the high-temperature pIRIR signals. Our result suggests that sensitivity correction could be problematic and special attention should be paid to sensitivity correction when using high temperature treatment in these protocols.

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